



# **Pulmonary Nocardiosis**

## **Similarity to Tuberculosis**

### **(A Bacteriological and Proteomics Study)**

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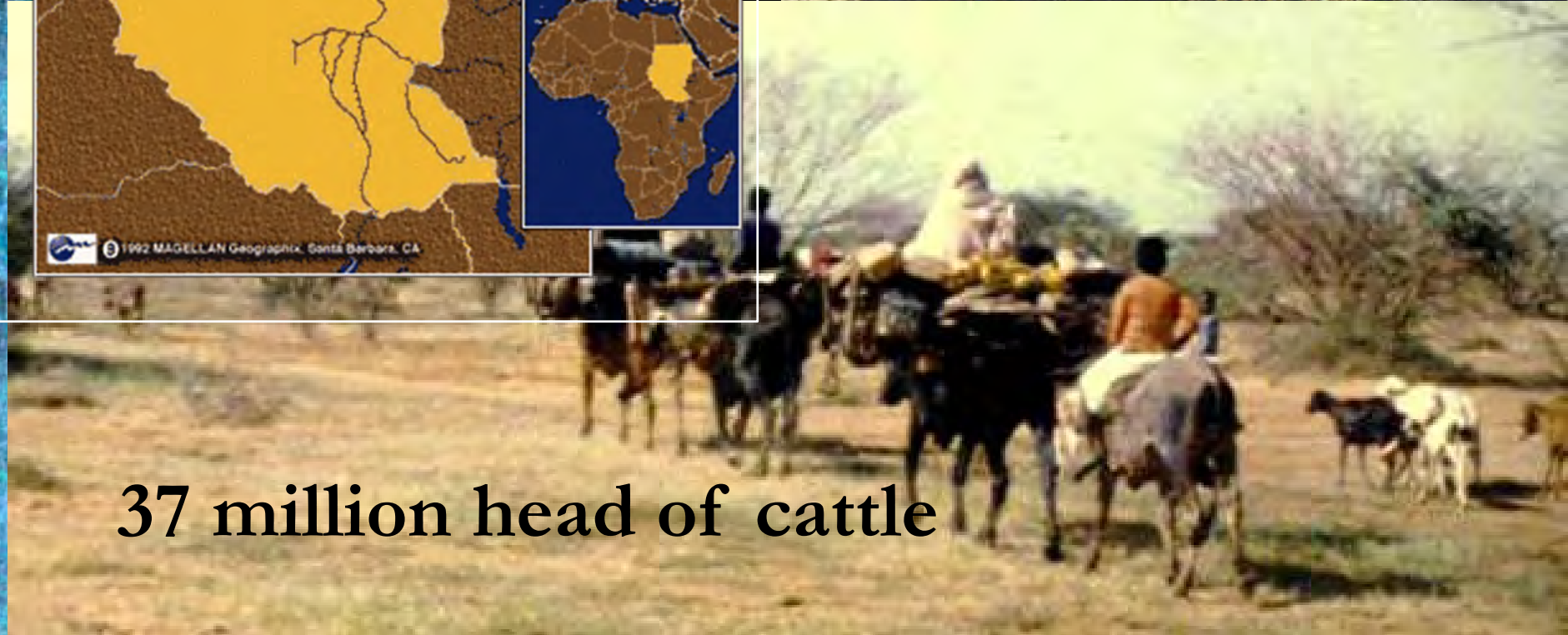
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# INTRODUCTION




38 million head of goats



37 million head of cattle

- **Nocardiae, are actinomycetes which are epidemiologically saprophytic**
- **They include species forming parasitic association with animals and plants**
- **They occur in a wide range of man made and natural habitat including activated sewage sludge, soil, water and tissues of plants and animals including human.**

- 
- **Nocardiosis, is an important cause of morbidity and mortality in patients with HIV and cancer (immunocompromised).**
  - **Nocardiosis is considered to be a late presenting community acquired infection, but there is growing evidence that the disease is transmissible.**



# Pulmonary Nocardiosis

- **Primary pulmonary nocardiosis may be sub clinical or pneumonic; it may be chronic or acute with secondary involvement of other organs, mainly the brain.**

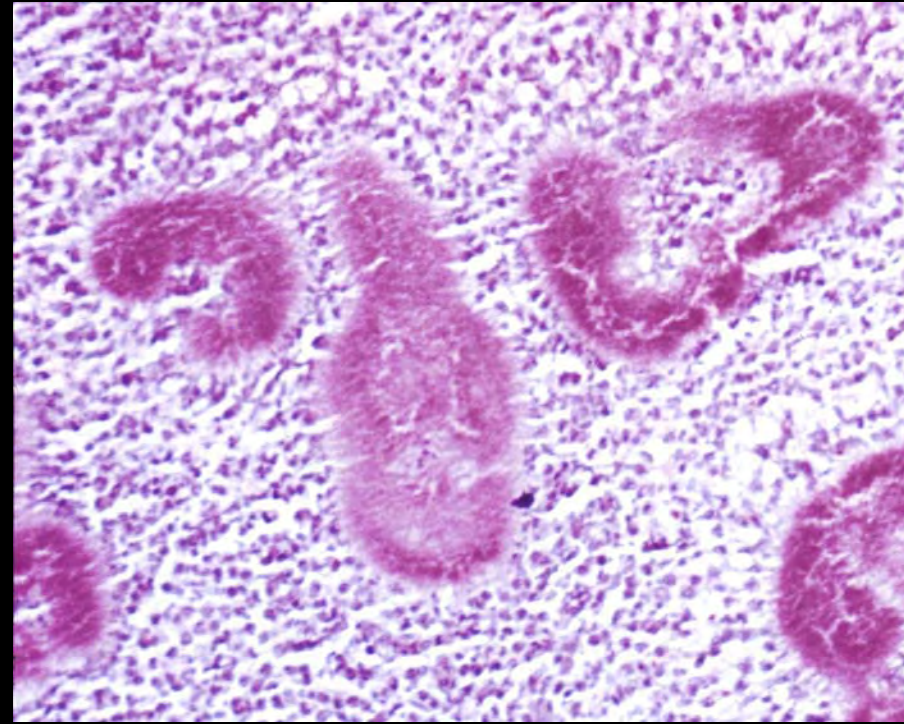
# Pulmonary Nocardiosis

- In non-tropical countries, most infections are caused by *N. asteroides*, *N. farcinica* and *N. nova*, relatively few by *N. brasiliensis*, *N. otitidiscaviarum* and *N. transvalenses*.

# Nocardia and Diseases



**Mastitis caused by *N. asteroides***



**Mycetoma produced by *Nocardia brasiliensis***

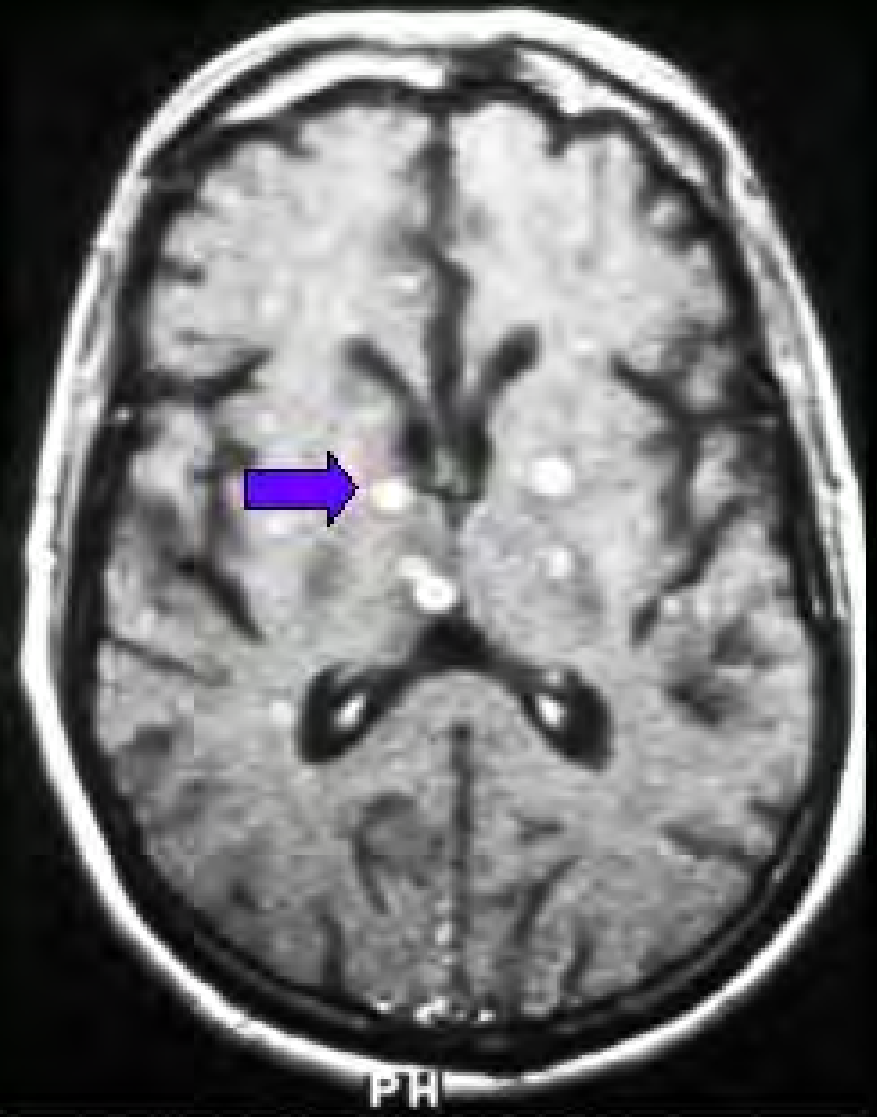
Dr. K. Salfelder



Dr. K. Salfelder

## CT Scan Image

Lesions caused by  
*Nocardia asteroides*  
infection in an HIV-  
positive individual



PH

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# ***Nocardia africana***

- ***Nocardia africana*, was isolated from sputum obtained from patients suffering from pulmonary infection admitted at Chest Unit of Khartoum Teaching Hospital in Sudan.**
- **The isolates showed morphological features of actinomycetes and they have assigned to the genus, *Nocardia*. According to their morphological criteria and chemical properties (Hamid *et al.*, 2001).**



# Proteomics

- **Recently, proteomics have been introduced as a new field of research for ultra study of protein structure and function**
- **Proteomics is the study of all proteins expressed by a genome. It involves the identification of proteins in the body and the determination of their role in physiological and pathophysiological functions.**

# Rationale

- **New strategies are required for diagnosis, prevention and therapy of tuberculosis as well as chest infections due to MOTT or nocardiae as it is misdiagnosed by physicians;**
- **An imminent example in Sudan was *Nocardia africana* which was thought to be multi-drug resistant *Mycobacterium tuberculosis*.**

# Objectives

## General Objectives:

To analyze all proteins expressed by the genome of *Nocardia africana* and *Nocardia farcinica* and to compare them to one of the most important pulmonary pathogens: *Mycobacterium tuberculosis*.

# Specific Objectives

- . To compare the protein compositions of *Nocardia africana* strains with *Nocardia farcinica* and *Nocardia asteroides*.
- . To compare the protein compositions of *Nocardia* spp. with *Mycobacterium tuberculosis*.



# **(A) Bacteriological Study**

# Methods

- Patients were examined for the presence of acid- fast bacilli.
- They were suspected to have tuberculosis infection according to the symptoms.

# Methods

- Thin bacterial smears were prepared and stained by Ziehl Neelsen (ZN) stain.
- Samples were then cultured on (L.J) medium
- Growth was monitored daily during the first week to observe the presence of rapid growers and then weekly up to the 8th week
- Phenotypic characterization was performed by using different biochemical tests.



# **(B) Proteomics Study**

**Test Strains**

**Protein Purification**

**Protein analysis**

# Preparation of Lysate

- *Nocardia species* were cultured on (GYEA) for 2 days and a lysate was prepared.
- Protein was purified and up-concentrated using the CleanUp Kit from BioRad

# Protein Purification

- **Two-dimensional gel electrophoresis was performed according to general protocols of BioRad with only slight modifications.**
- **17 cm pH3-10 IPG strips were actively rehydrated and gels were stained with Coomassie Brilliant Blue and subsequently analyzed with PDQuest 6.2 (BioRad).**

# Protein analysis


- Prominent spots were excised and tryptically digested in the gel .
- For matrix-assisted laser desorption/ionization (MALDI)-MS, samples were purified using ZipTips C18
- Peptide maps were generated with MALDI microMX instrument .
- $\alpha$ -cyano-4-hydroxycinnamic acid was used as matrix and 1  $\mu$ l was applied.


# Protein analysis

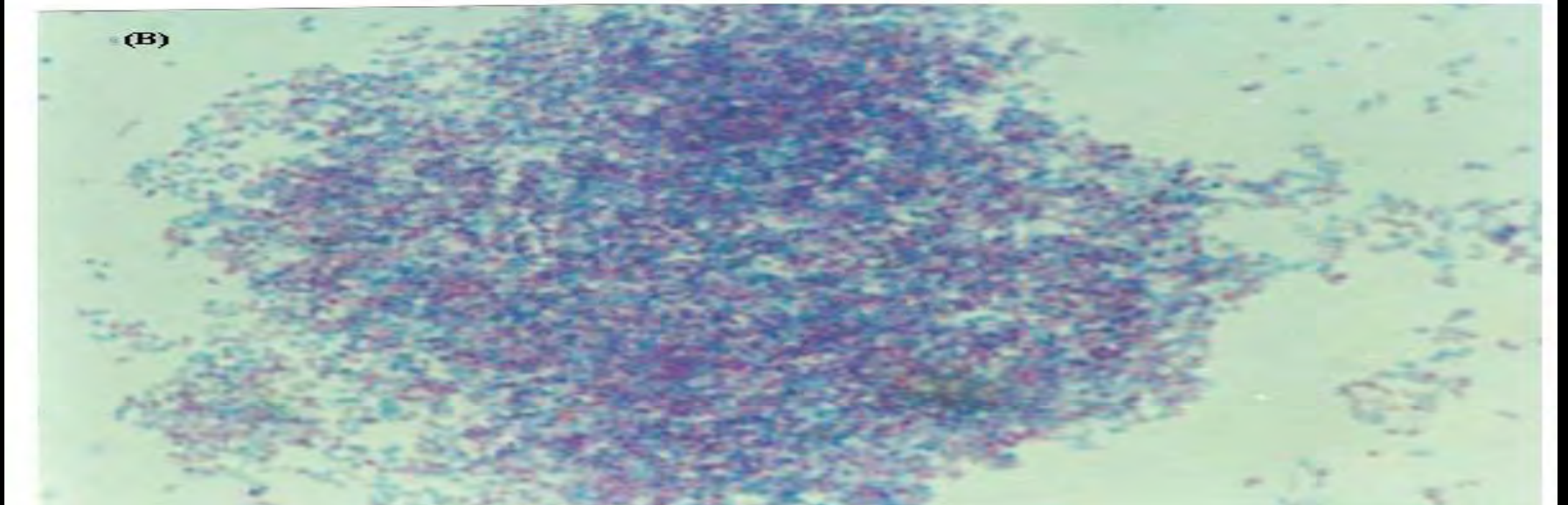
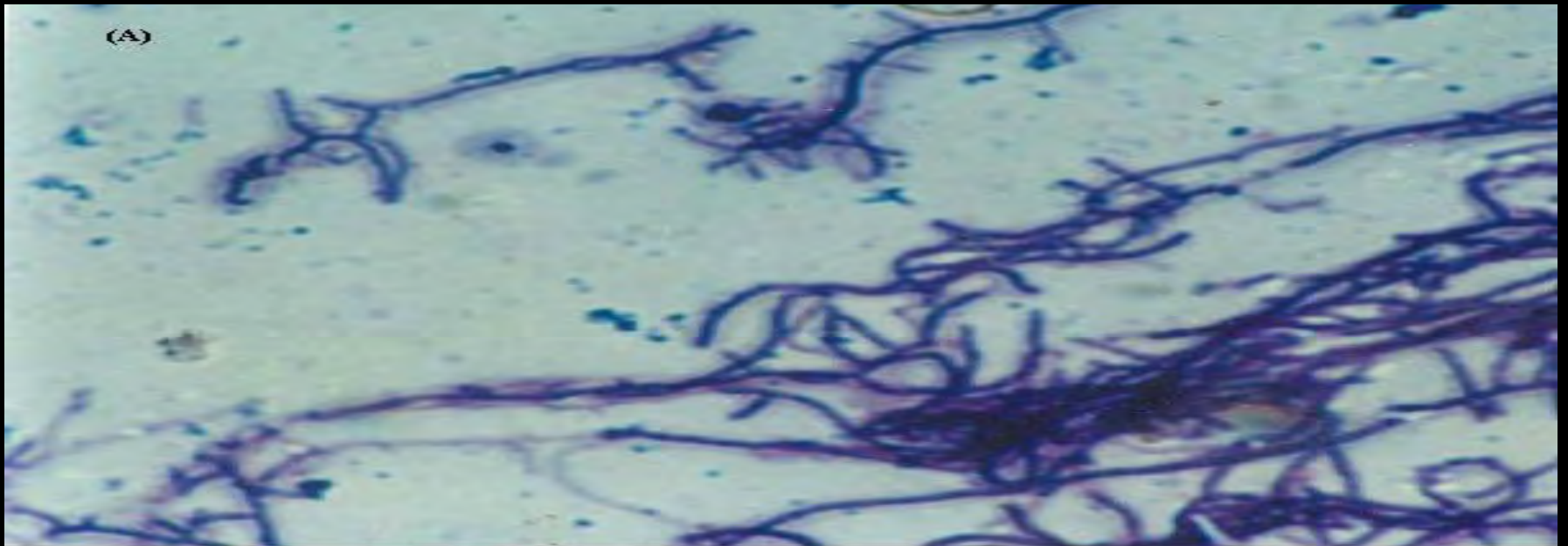
- Database queries were carried out using Mascot in-house with species limitation on bacteria.
- All data can be obtained from the NoDaMS site (Nocardia database on Mass spectrometry analyses) on the following web address: <http://ifg-izkf.uni-muenster.de/proteomik/>.

# RESULTS AND DISCUSSION

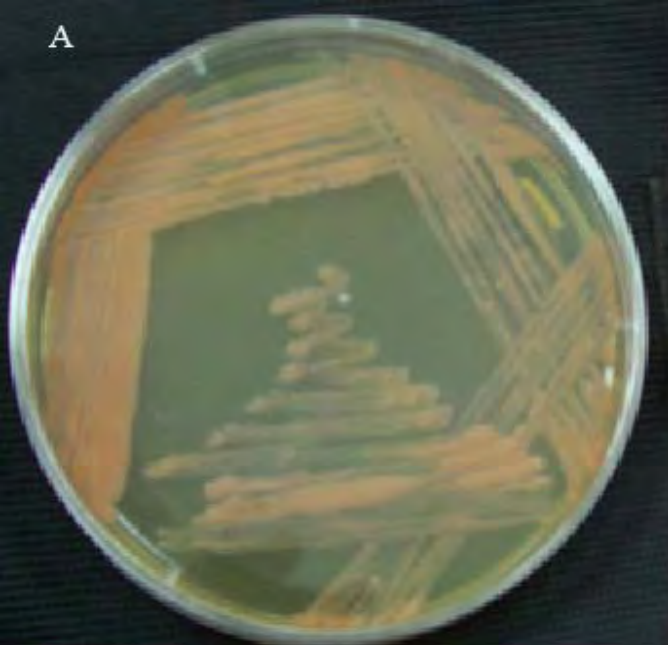
# BACTERIOLOGICAL RESULTS

- 
- The colonial morphology of 319 (97%) appeared as rough, friable, warty, granular and grey in color with irregular margins and showed the appearance of AFB when stained again (indirect smear).
  - The 319 isolates were initially identified as members of the *M. tuberculosis* complex.

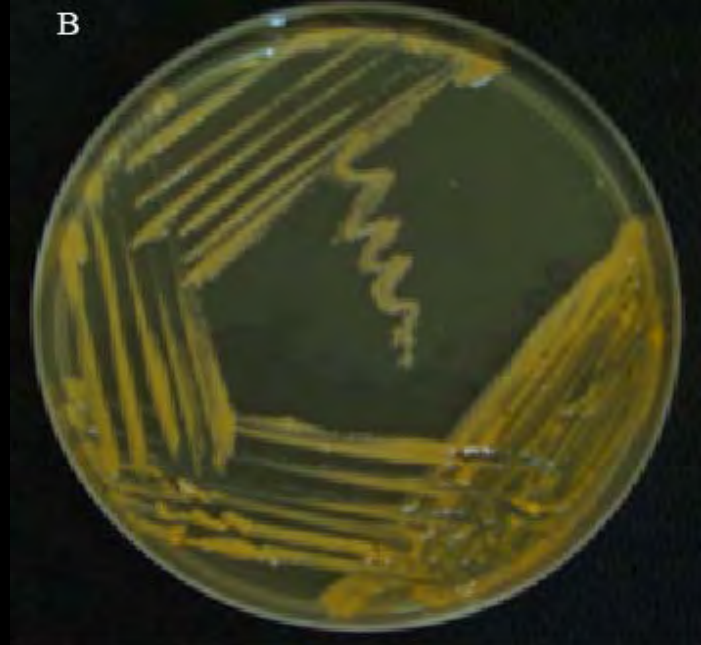
- 
- **Ten (3%) of the LJ slopes revealed the growth of small orange filamentous colonies, (within 2-3 days) which were tentatively considered to be nocardiae. Selected biochemical tests were performed to confirm this findings**
  - **The 10 nocardia isolates, were then designated SD1001, SD1002, SD1003, SD1004, SD1005, SD1006, SD1007, SD1008, SD1009 and SD10010**



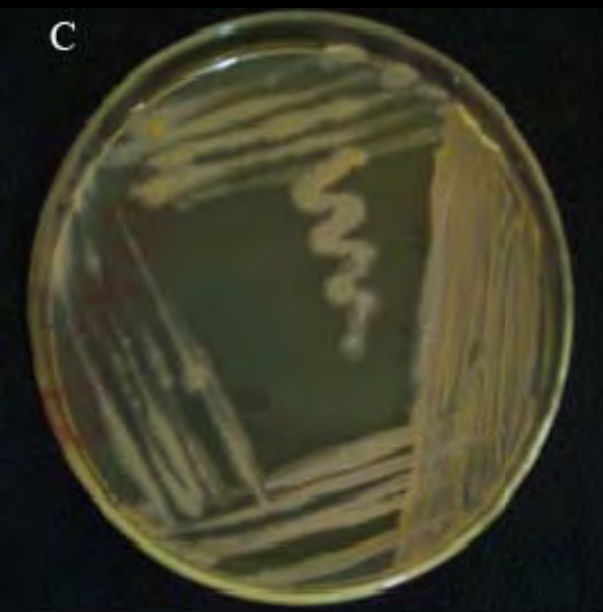
A



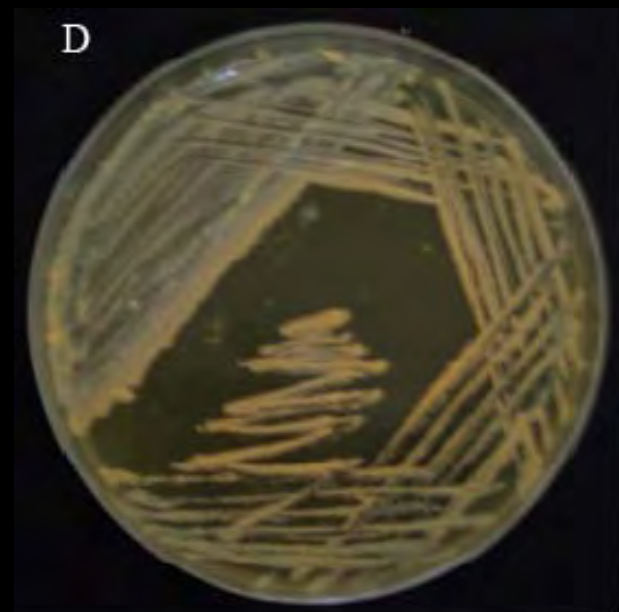
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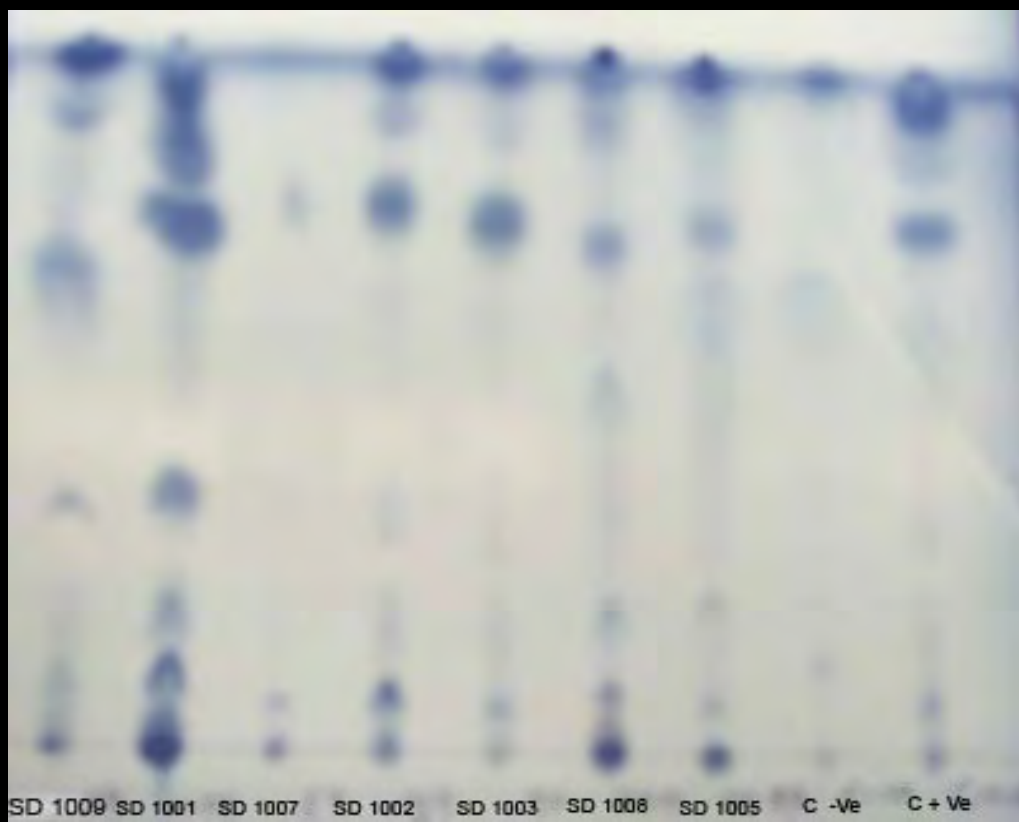


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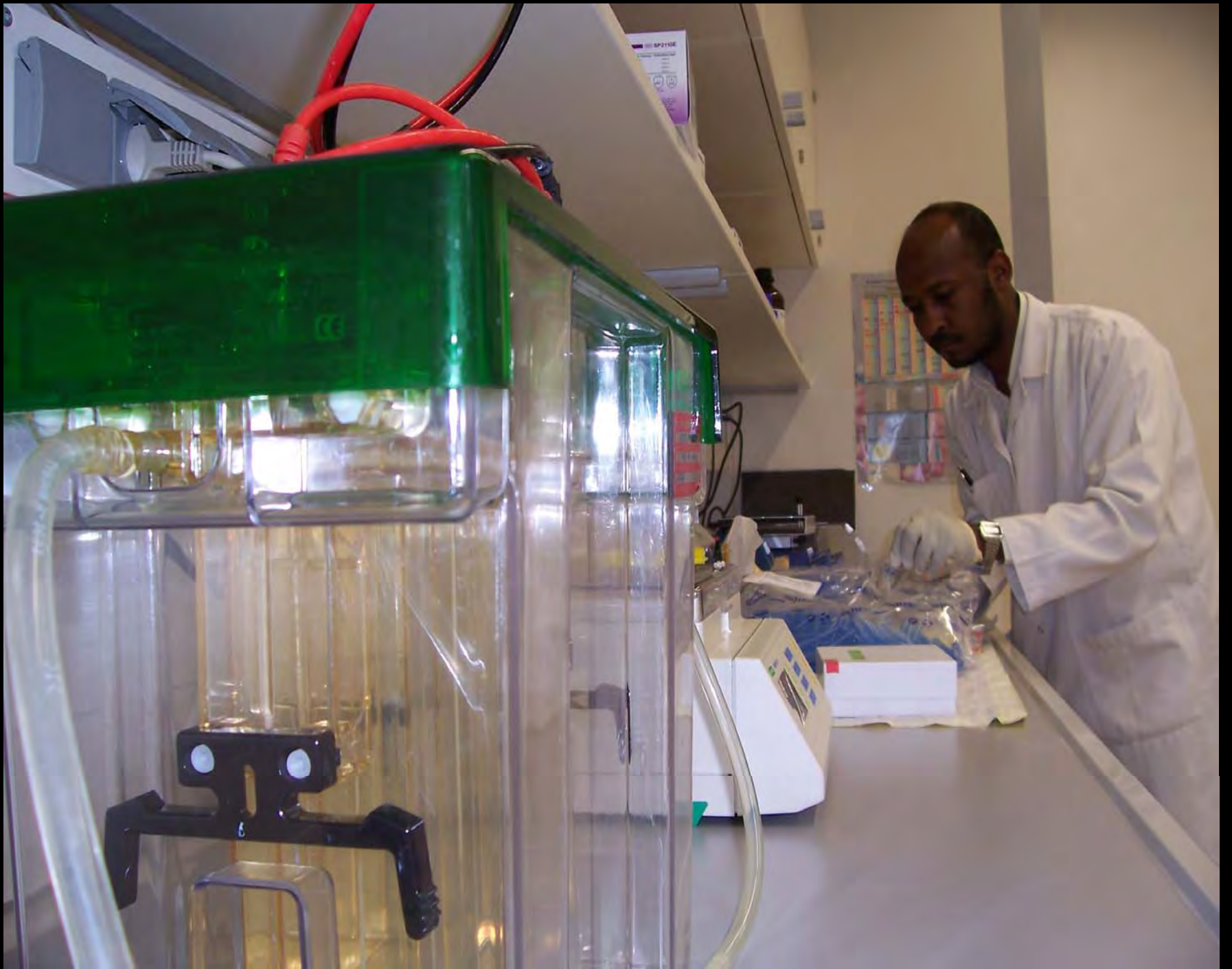
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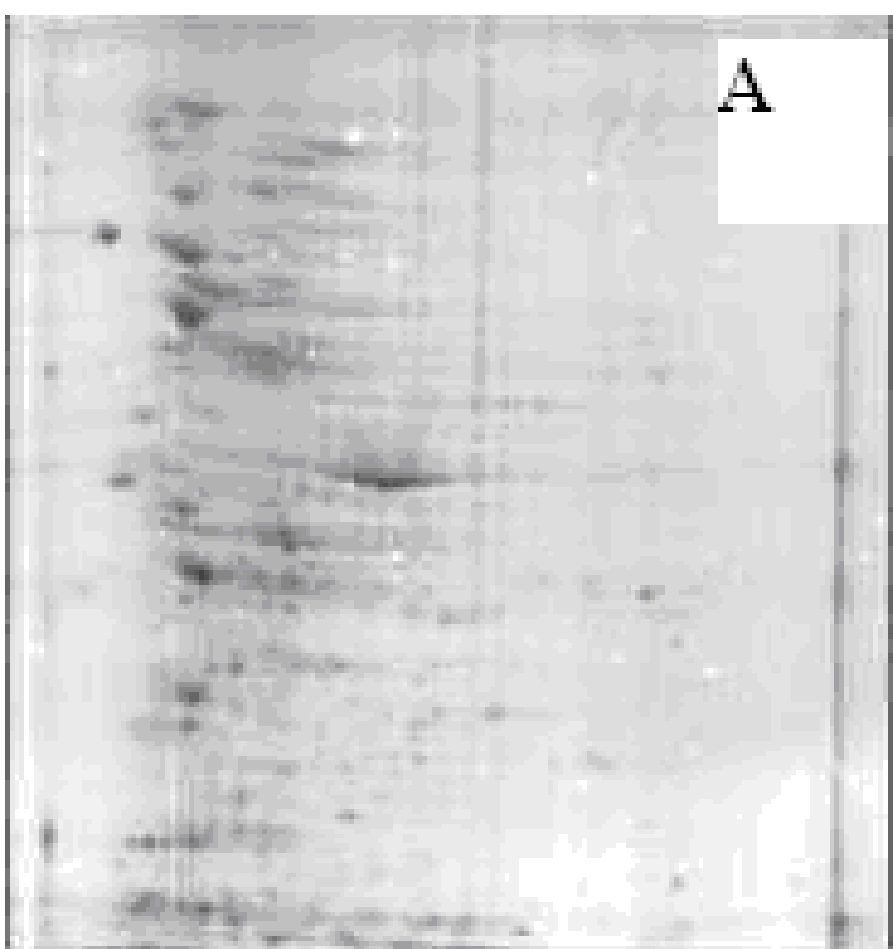


# PROTEOMICS RESULTS

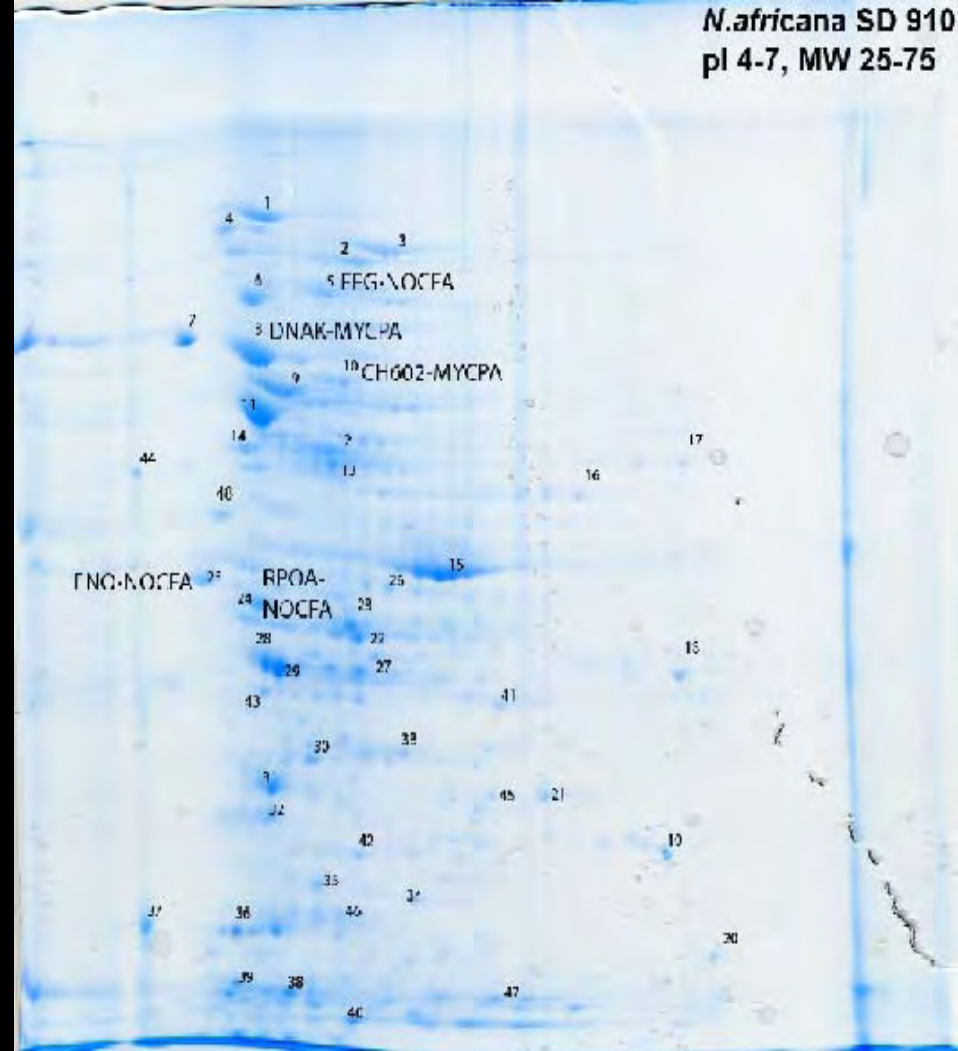
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- **Two Dimensional Poly Acrylamide Gel Electrophoresis (2D-PAGE) using pH strips 3-10 revealed that the soluble proteins were visible in a much smaller pI range. All strains exhibited similar protein distributions. A similarity analysis revealed that mycobacterium sequences are of high relevance for the investigated strains.**





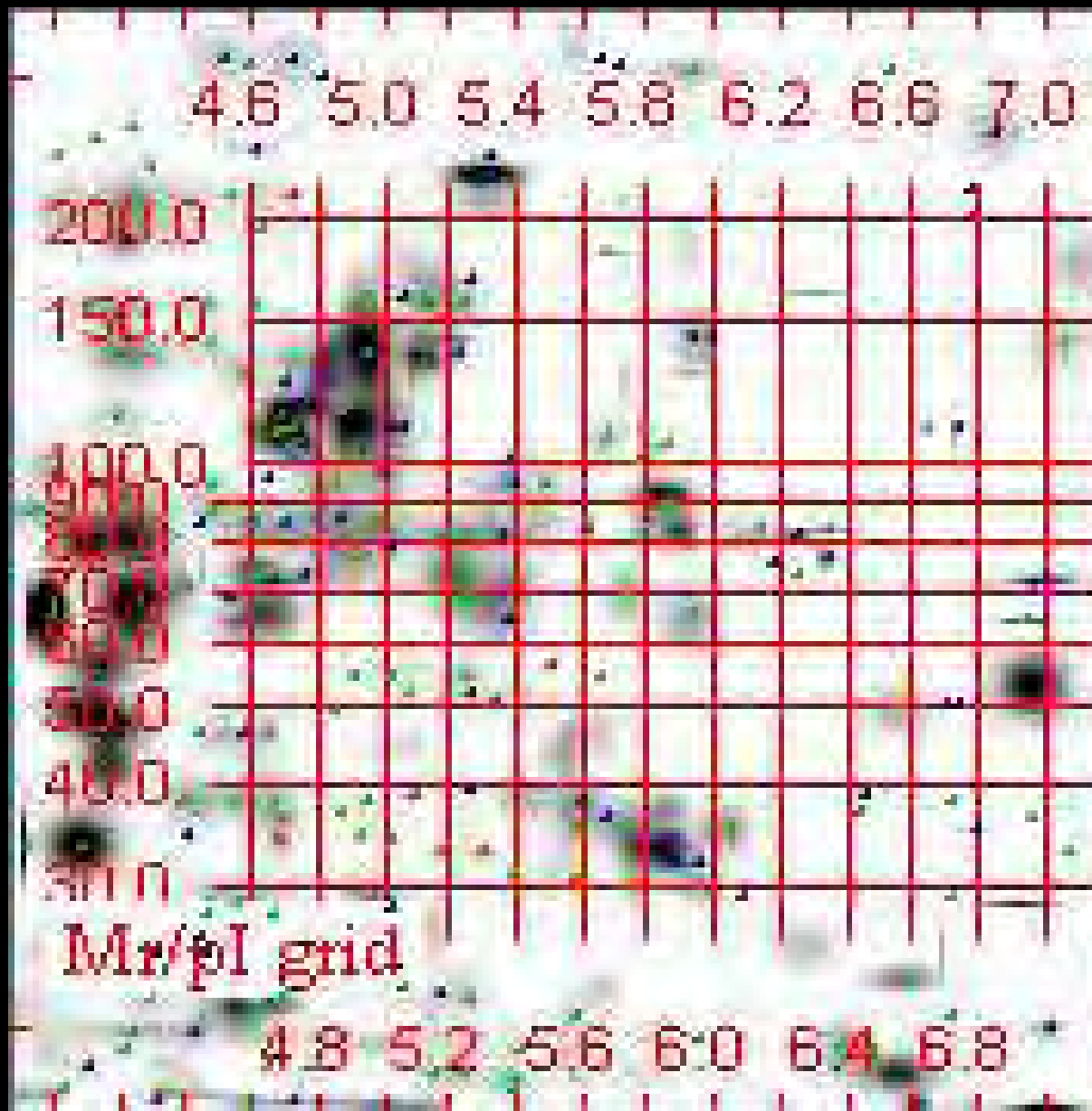


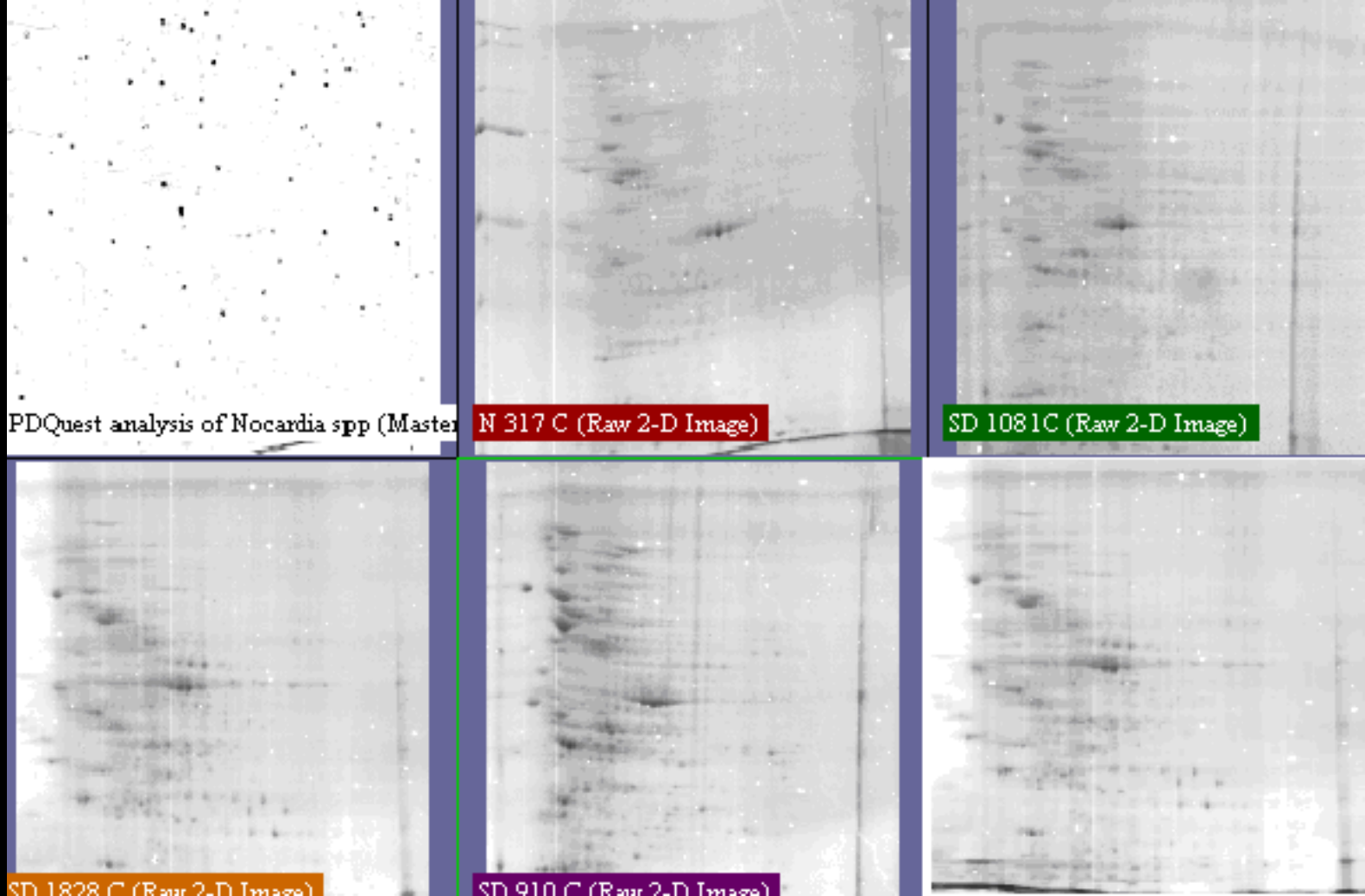
SD 910



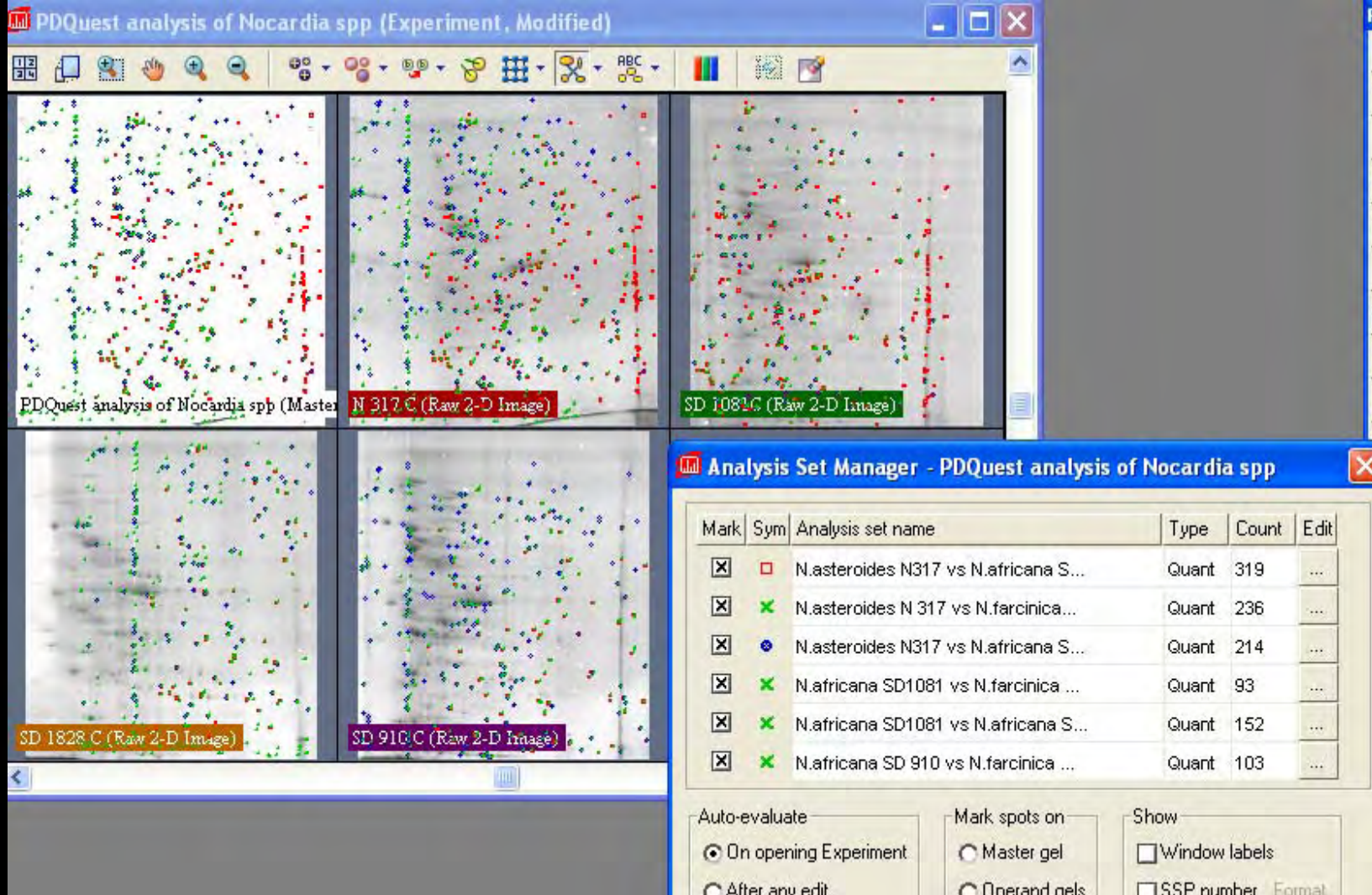
**Gel images of *N. africana*  
SD910**



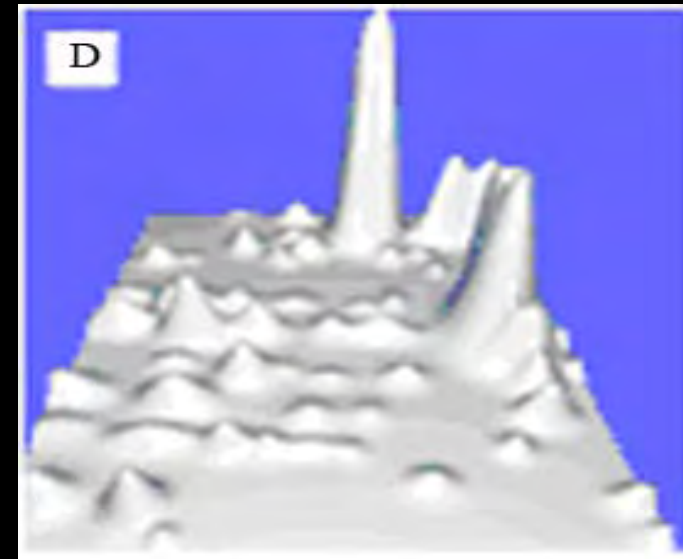
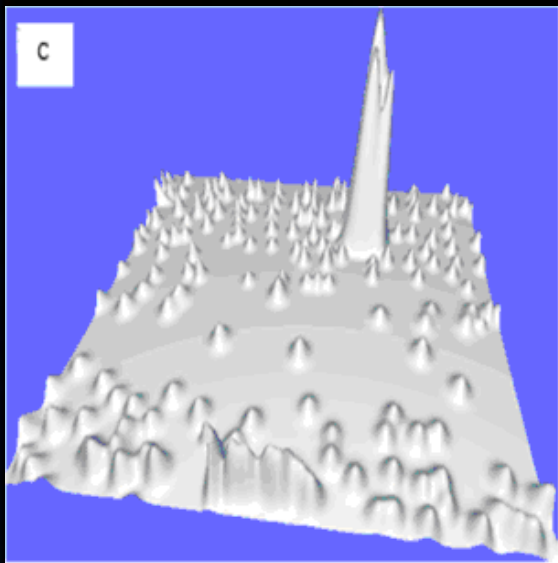
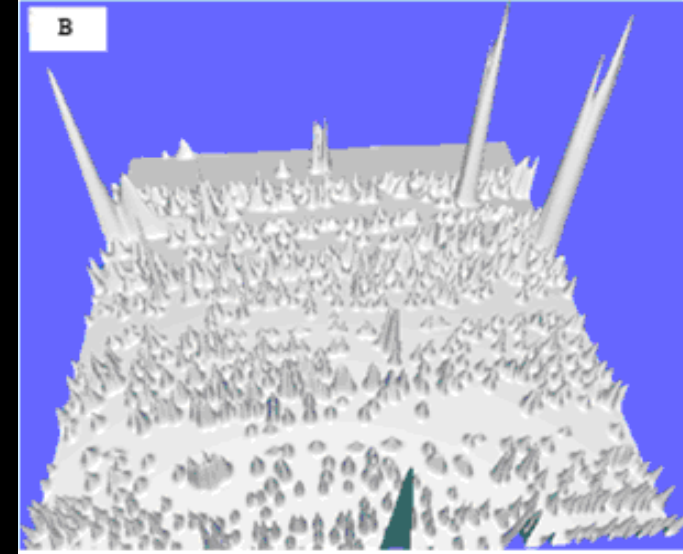
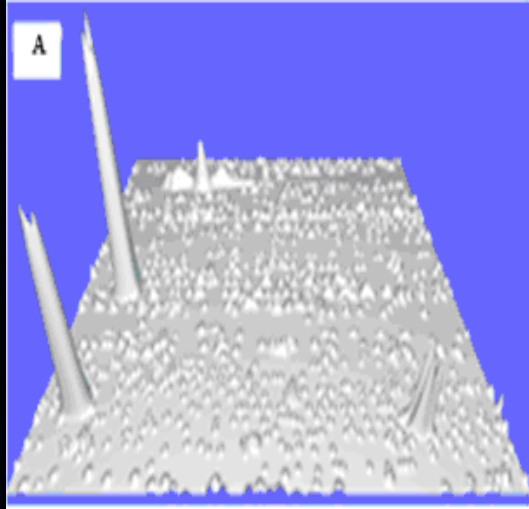




Comparison of filtered Gel images of *N. asteroides* N317, *Nocardia. sp.* N 1086, *N. farcinica* SD1828, *N. africana* SD910 and *N. africana* SD925 with master gel as analyzed with PDQuest.



Overlay of gels of analyzed with PDQuest.



**3 D view of Spots in the central regions of the gels**





Finger print of common peptides detected Elongation factors  
of *N. africana* SD910

spot5 assigned to elongation factor G (EF-G; Q5YPG3;  
EFG\_NOCFA, 76971).

| <b>MW</b> | <b>Start</b> | <b>End</b> | <b>Peptide</b>                |
|-----------|--------------|------------|-------------------------------|
| 1014.57   | 320          | 328        | K.IAVHPFFGK.L                 |
| 1090.54   | 124          | 132        | R.QADKYDVPR.I                 |
| 1216.65   | 32           | 41         | R.ILFYTG VNYK.I               |
| 1429.77   | 475          | 487        | K.VEANVGK PQVAYR.E            |
| 1442.68   | 112          | 123        | K.EGV EPQSEQVWR.Q             |
| 1772.91   | 142          | 156        | K.LGADFYFTVQT IKDR.L          |
| 1779.92   | 651          | 666        | K.ALVPLSEMFGYIGDLR.S          |
| 1795.92   | 651          | 666        | K.ALVPLSEMFGYIGDLR.S          |
| 1833.97   | 472          | 487        | R.EFKVEANVGK PQVAYR.E         |
| 1893.98   | 233          | 249        | K.FFGGEELTIDEIKG AIR.K        |
| 2022.08   | 233          | 250        | K.FFGGEELTIDEIKG AIRK.M       |
| 2614.30   | 100          | 123        | R.VLDGAVAVFDGKEGV EPQSEQVWR.Q |



**Searching homology for the  
isolated proteins BLAST2  
search results**

**([http://www.expasy.org/cgi-  
bin/blast.pl](http://www.expasy.org/cgi-bin/blast.pl))**



# Elongation factor Tu of *Mycobacterium leprae* EFTU\_MYCLE (P30768)

SwissProt Accession #

Organism

Q73SD1

*Mycobacterium paratuberculosis*

P0A558

*Mycobacterium tuberculosis*

P0A559

*Mycobacterium bovis*

Q1BDD3

*Mycobacterium* sp. (strain MCS)

Q1T9Z3

*Mycobacterium* sp. KMS

Q1TV69

*Mycobacterium* sp. JLS

Q262Y2

*Mycobacterium vanbaalenii* PYR-1

Q27DQ3

*Mycobacterium flavescence*

Q5YPG4

*Nocardia farcinica*

**Chaperone protein dnaK, HSP70 of *Mycobacterium paratuberculosis*  
DNAK\_MYCPA (Q00488)**

| <b>Description</b>     | <b>SwissProt Accession<br/>#</b> | <b>Organism</b>                                 |
|------------------------|----------------------------------|---|
| <b>DNAK_MYC<br/>LE</b> | <b>P19993</b>                    | <b><i>Mycobacterium leprae</i></b>              |
| <b>DNAK_MYC<br/>TU</b> | <b>P0A5B9</b>                    | <b><i>Mycobacterium tuberculosis</i></b>        |
| <b>DNAK_MYC<br/>BO</b> | <b>P0A5C0</b>                    | <b><i>Mycobacterium bovis</i></b>               |
| <b>_MYCVN</b>          | <b>Q25TL7</b>                    | <b><i>Mycobacterium vanbaalenii</i> PYR-1</b>   |
| <b>_MYCFV</b>          | <b>Q27CE8</b>                    | <b><i>Mycobacterium flavescence</i> PYR-GCK</b> |
| <b>_MYCSS</b>          | <b>Q1BEV1</b>                    | <b><i>Mycobacterium</i> sp. strain MCS</b>      |
| <b>_9MYCO</b>          | <b>Q1TWF2</b>                    | <b><i>Mycobacterium</i> sp. JLS</b>             |
| <b>_9MYCO</b>          | <b>Q1TDC6</b>                    | <b><i>Mycobacterium</i> sp. KMS</b>             |
| <b>DNAK_NOCF<br/>A</b> | <b>Q5YNI0</b>                    | <b><i>Nocardia farcinica</i></b>                |



## 60 kDa chaperonin 2 of *N. farcinica* CH602\_NOCF A (Q9AFA6)

| <b>Description</b>      | <b>SwissProt<br/>Accession #</b> | <b>Organism</b>  |
|-------------------------|----------------------------------|--|
| <b>CH60_CO<br/>REQ</b>  | <b>Q93QI2</b>                    | <i>Corynebacterium equii</i> ( <i>Rhodococcus<br/>equi</i> ) |
| <b>_RHOSR</b>           | <b>Q0SET3</b>                    | <i>Rhodococcus</i> sp. strain <i>RHA1</i>                    |
| <b>_MYCFV</b>           | <b>Q27CR6</b>                    | <i>Mycobacterium flavescence</i> <i>PYR-GCK</i>              |
| <b>_MYCSS</b>           | <b>Q1BEF6</b>                    | <i>Mycobacterium</i> sp. strain <i>MCS</i>                   |
| <b>_9MYCO</b>           | <b>Q1TWU6</b>                    | <i>Mycobacterium</i> sp. <i>JLS</i>                          |
| <b>_MYCVN</b>           | <b>Q267S7</b>                    | <i>Mycobacterium vanbaalenii</i> <i>PYR-1</i>                |
| <b>_9MYCO</b>           | <b>Q1TDR8</b>                    | <i>Mycobacterium</i> sp. <i>KMS</i>                          |
| <b>_9MYCO</b>           | <b>Q8GAR8</b>                    | <i>Mycobacterium</i> sp. <i>185-409</i>                      |
| <b>CH60_NO<br/>CAS</b>  | <b>Q9AFC5</b>                    | <i>Nocardia asteroides</i>                                   |
| <b>_MYCMR</b>           | <b>Q8G8X0</b>                    | <i>Mycobacterium marinum</i>                                 |
| <b>CH602_M<br/>YCPA</b> | <b>P42384</b>                    | <i>Mycobacterium paratuberculosis</i>                        |

## Enolase of *N. farcinica* ENO\_NOCFA (Q5YQ30)

| Description | SwissProt<br>Accession # | Organism                                    |
|-------------|--------------------------|---|
| _RHOSR      | Q0S4I1                   | <i>Rhodococcus sp. strain<br/>RHA1</i>      |
| _MYCSS      | Q1B439                   | <i>Mycobacterium sp. strain<br/>MCS</i>     |
| _9MYCO      | Q1TAN9                   | <i>Mycobacterium sp. KMS</i>                |
| _9MYCO      | Q1TX18                   | <i>Mycobacterium sp. JLS</i>                |
| _MYCFV      | Q275H2                   | <i>Mycobacterium flavescens<br/>PYR-GCK</i> |
| ENO_MYCPA   | Q741U7                   | <i>Mycobacterium vanbaalenii<br/>PYR-1</i>  |
| _MYCVN      | Q267Y1                   | <i>Mycobacterium<br/>tuberculosis</i>       |
| ENO_MYCBO   | Q7U0U6                   | <i>Mycobacterium bovis</i>                  |
| ENO_MYCTU   | P96377                   | <i>Mycobacterium<br/>tuberculosis</i>       |
| ENO_MYCLE   | Q9CD42                   | <i>Mycobacterium leprae</i>                 |

# Transkriptase alpha chain of *N. farcinica* RPOA\_NOCFA (Q5Z1K9)

| Description | SwissProt Accession # | Organism                                |
|-------------|-----------------------|---|
| _RHOSR      | Q0S3E7                | <i>Rhodococcus</i> sp. strain RHA1      |
| _MYCSS      | Q1BD08                | <i>Mycobacterium</i> sp. strain MCS     |
| _9MYCO      | Q1T8C7                | <i>Mycobacterium</i> sp. KMS            |
| _9MYCO      | Q1TR19                | <i>Mycobacterium</i> sp. JLS            |
| _MYCFV      | Q27E10                | <i>Mycobacterium flavescens</i> PYR-GCK |
| _MYCVN      | Q263M4                | <i>Mycobacterium vanbaalenii</i> PYR-1  |
| RPOA_MYCTU  | P66701                | <i>Mycobacterium tuberculosis</i>       |
| RPOA_MYCBO  | P66702                | <i>Mycobacterium bovis</i>              |
| RPOA_MYCPA  | Q73S43                | <i>Mycobacterium paratuberculosis</i>   |
| RPOA_MYCLE  | Q9X798                | <i>Mycobacterium leprae</i>             |

# CONCLUSIONS


- **Similarity in clinical manifestations between pulmonary nocardiosis and pulmonary tuberculosis and ZN results needs further investigations by different methods in order to differentiate between the different causative agents and to describe the correct treatment regimen.**

# CONCLUSIONS

- A proteomics study was found useful, a protocol for cell lysis was successfully conducted on nocardia cells.
- A comparative analysis using PDQuest software, showed the similarities between different nocardia species investigated as presented by 2D gels.
- Peptides finger print using MALDI- MS. of different proteins confirmed the existence of various vital proteins in all *Nocardia* isolates.

# CONCLUSIONS

- **MALDI- MS. analysis showed that there were many proteins which were restricted to a certain *Nocardia* species comparing with others, thus this or that protein can be used to distinguish between different *Nocardia* spp.**
- **Homology of the isolated proteins revealed that *Mycobacterium* and to a lesser extent *Rhodococcus* sequences are of high relevance for the five investigated nocardia strains.**
- **For a successful proteomics protocol, the high quality pure protein preparation is a critical point, which is difficult to achieve in the case of *Nocardia* as it is rich with mycolic acids.**


- 
- **Thus, more additional steps for protein purification and concentration were necessary. High specialized laboratory facilities are needed as well.**
  - **Moreover, the proteome data concerning Nocardiae in the data bases is so limited, which makes the alignment of isolated proteins a difficult task.**



# PROSPECTIVE WORK

# FUTURE WORK

- **A DIGE (Differential Gel Electrophoresis) based approach is recommended for further improvements, because classical 2-D gel electrophoresis has a considerable degree of gel-to-gel variations that can be minimized by DIGE.**
- **A large body of MS data is available in the NoDaMS database which was created depending on the results obtained from this study, to share peak lists for future investigations using advanced protein databases**

- 
- **Further proteomics studies are essential to establish the complete proteome of *Nocardia africana* as well as other *Nocardia* spp. For this approach, different types of culture media and variable condition are necessary to investigate the possibility of having other secretory and/or excretory proteins**



# AKNOLEDGMENTS

- **SUST**
- **DAAD**
- **ICGEB**
- **Mr. Adil Mahgoub**
- **Dr. Kanory Rao**
- **Staff of TB Reference lab**
- **Staff of Abu-Anga Hospital**
- **Staff of El Shaab Teaching Hospital**



**THANK YOU**